



The
Patent
Office

PCT/GB 93 / 02367

REC'D 20 JAN 1994

WIPO PCT

The Patent Office
Cardiff Road
Newport
Gwent
NP9 1RH

08 / 436265

I, the undersigned, being an officer duly authorised in accordance with Section 62(3) of the Patents and Designs Act 1907, to sign and issue certificates on behalf of the Comptroller-General, hereby certify that annexed hereto is a true copy of the documents as originally filed in connection with the Patent application identified therein.

In accordance with the Patents (Companies Re-registration) Rules 1982, if a company named in this certificate and any accompanying documents has re-registered under the Companies Act 1980 with the same name as that with which it was registered immediately before re-registration save for the substitution as, or the inclusion as, the last part of the name of the words "public limited company" or their equivalents in Welsh, references to the name of the company in this certificate and any accompanying documents shall be treated as references to the name with which it is so re-registered.

In accordance with the rules, the words "public limited company" may be replaced by p.l.c., plc, P.L.C. or PLC.

Re-registration under the Companies Act does not constitute a new legal entity but merely subjects the company to certain additional company law rules.

Signed

M. Russell

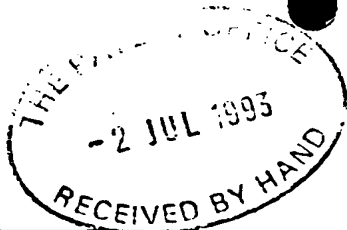
Dated

6th January 1994

CO

PRIORITY DOCUMENT

For official use



9313763.6

Your reference

70/4201/01

02 JUL 1993

Notes

Please type, or write in dark ink using CAPITAL letters. A prescribed fee is payable for a request for grant of a patent. For details, please contact the Patent Office (telephone 071-438 4700).

Rule 16 of the Patents Rules 1990 is the main rule governing the completion and filing of this form

Do not give trading styles, for example, 'Trading as XYZ company', nationality or former names, for example, 'formerly (known as) ABC Ltd' as these are not required.

Warning

After an application for a Patent has been filed, the Comptroller of the Patent Office will consider whether publication or communication of the invention should be prohibited or restricted under Section 22 of the Patents Act 1977 and will inform the applicant if such prohibition or restriction is necessary. Applicants resident in the United Kingdom are also reminded that under Section 23, applications may not be filed abroad without written permission unless an application has been filed not less than 6 weeks previously in the United Kingdom for a patent for the same invention and either no direction prohibiting publication or communication has been given, or any such direction revoked.

The Patent Office

Request for grant of a Patent Form 1/77

Patents Act 1977

1 Title of invention

1 Please give the title of the invention **PROTEIN KINASES**

2 Applicant's details

☐ **First or only applicant**

2a If you are applying as a corporate body please give:

Corporate name **Ludwig Institute for Cancer Research**

Country (and State of incorporation, if appropriate) **United Kingdom**

2b If you are applying as an individual or one of a partnership please give in full:

Surname

Forenames

2c In all cases, please give the following details:

Address **St. Mary's Hospital Medical School
Norfolk Place
Paddington
London**

UK postcode (if applicable) **W2 1PG**

Country **England**

ADP number (if known)

2e and 2f: If there are further applicants please provide details on a separate sheet of paper.

☐ **Second applicant (if any)**

2d If you are applying as a corporate body please give:
Corporate name

Country (and State
of incorporation, if
appropriate)

2e If you are applying as an individual or one of a partnership please give in full:

Surname

Forenames

2f In all cases, please give the following details:

Address

UK postcode
(if applicable)

Country
ADP number
(if known)

3 An address for service in the United Kingdom must be supplied

Please mark correct box

3 Address for service details

3a Have you appointed an agent to deal with your application?

Yes ☒ No ☐ → go to 3b

↓
please give details below

Agent's name

Gill Jennings and Every

Agent's address

Broadgate House
7 Eldon Street
London

Postcode

EC2M 7LH

Agent's ADP
number

745002

3b: If you have appointed an agent, all correspondence concerning your application will be sent to the agent's United Kingdom address.

3b If you have not appointed an agent please give a name and address in the United Kingdom to which all correspondence will be sent:

Name

Address

Postcode

ADP number
(if known)

Daytime telephone
number (if available)

4 Reference number

4 Agent's or
applicant's reference 70/4201/01
number (if applicable)

5 Claiming an earlier application date

5 Are you claiming that this application be treated as having been filed on the date of filing of an earlier application?

Yes ☐ No ☒ ⇒ go to 6

↓
please give details below

☐ number of earlier
application or patent
number

☐ filing date

(day month year)

☐ and the Section of the Patents Act 1977 under which you are claiming:

15(4) (Divisional) ☐ 8(3) ☐ 12(6) ☐ 37(4) ☐

6 Declaration of priority

6 If you are declaring priority from previous application(s), please give:

Country of filing	Priority application number (if known)	Filing date (day, month, year)
-------------------	---	-----------------------------------

--	--	--

Please mark correct box

Please mark correct box

6 If you are declaring priority from a PCT application please enter 'PCT' as the country and enter the country code (for example, GB) as part of the application number.

Please give the date in all number format, for example, 31/05/90 for 31 May 1990.

7 The answer must be 'No' if:
any applicant is not an inventor

- there is an inventor who is not an applicant, or
- any applicant is a corporate body.

8 Please supply duplicates of claim(s), abstract, description and drawing(s).

Please mark correct box(es)

9 You or your appointed agent (see Rule 90 of the Patents Rules 1990) must sign this request.

Please sign here ⇒

A completed fee sheet should preferably accompany the fee.

7 Inventorship

7 Are you (the applicant or applicants) the sole inventor or the joint inventors?

Please mark correct box

Yes ☐ No ☒

A Statement of Inventorship on Patents

Form 7/77 will need to be filed (see Rule 15)

8 Checklist

8a Please fill in the number of sheets for each of the following types of document contained in this application.

Continuation sheets for this Patents Form 1/77

0

Claim(s)

0

Description

7

Abstract

0

Drawing(s)

5

8b Which of the following documents also accompanies the application?

Priority documents (please state how many)

0

Translation(s) of Priority documents (please state how many)

0

Patents Form 7/77 - Statement of Inventorship and Right to Grant (please state how many)

0

Patents Form 9/77 - Preliminary Examination/Search

Patents Form 10/77 - Request for Substantive Examination

9 Request

I/We request the grant of a patent on the basis of this application.

For the Applicant

GILL JENNINGS & EVERY

Signed

R. E. Perry

Date 02 . 07 . 93

(day month year)

Please return the completed form, attachments and duplicates where requested, together with the prescribed fee to either:

☐ The Comptroller
The Patent Office
Cardiff Road
Newport
Gwent
NP9 1RH

or

☐ The Comptroller
The Patent Office
25 Southampton Buildings
London
WC2A 1AY

PROTEIN KINASES

This invention is related to the one or more inventions described in British Patent Applications Nos. 9224057.1, filed 17th November 1992, 9304677.9 and 9304680.3, both filed 8th March 1993, and 9311047.6, filed 28th May 1993. In particular, this invention relates to nucleotides, proteins obtained by expression therefrom and antibodies raised to peptides derived from the sequence, e.g. by the means described in the Application No. 9304680.3.

Summary: Mouse cDNA clones encoding putative serine/threonine kinase receptors denoted activin receptor-like kinase (ALK)-3 and -6 were obtained from a 12 day mouse embryo cDNA library. The cDNA clones for ALK-3 and -6 encode complete proteins of 532 and 502 amino acids, respectively. They have hydrophilic cysteine-rich ligand-binding domains, followed by single hydrophobic transmembrane regions and C-terminal intracellular portions containing putative serine/threonine kinase domains. The amino acid sequences of ALK-3 and -6 are very similar with 59% and 85% sequence identities in the extracellular cysteine-rich domains and intracellular serine/threonine kinase domains, respectively. Expression of mRNA for mouse ALK-3 was observed in the spleen, whereas ALK-6 mRNA was found in the brain. These results suggest that ALK-3 and -6 bind closely related ligands, but that they have different functional roles *in vivo*.

Transforming growth factor- β (TGF- β) is a family of multifunctional proteins, including TGF- β 1, - β 2, and - β 3 in mammals. They regulate the growth and differentiation of many different cells, and stimulate the production of extracellular matrix proteins (1). TGF- β s belong to a larger superfamily of structurally related proteins, which includes activins and inhibins, Müllerian inhibiting substance, and bone morphogenetic proteins (BMPs) or osteogenic proteins (2, 3). Proteins in the TGF- β superfamily regulate the cellular proliferation and differentiation, and play important roles at the different stages during the development.

TGF- β s exert their effects through binding to specific cell surface receptors (4, 5). Although TGF- β receptors of several different size classes have been reported, the type I (53 kDa) and type II (70 kDa) receptors have been shown to be most important for signal transduction (6, 7). TGF- β receptors type I and type II are indispensable for signal transduction, and they have been shown to form a heteromeric complex on the cell surface (8). In analogy with the TGF- β receptor system, classes of receptors of similar sizes, i.e. receptors type I (50-60 kDa) and type II (70-80 kDa), have been reported also for activin A and BMP-4 (9, 10).

Molecular cloning of the type II receptors for activin (ActR-II and ActR-IIB) (11-13) and for TGF- β (T β R-II) (14) revealed that they have sequence homology with each other and with a previously identified *C. elegans* gene product, Daf-1 (15). These

molecules have putative serine/threonine kinase domains in their intracellular portions, which suggests that phosphorylation on serine/threonine residues is involved in signal transduction. Using a polymerase chain reaction (PCR)-based strategy, we have recently obtained human cDNA clones encoding novel putative serine/threonine kinase receptors, denoted activin receptor-like kinases (ALK)-1 to -5 (16 and Franzén et al., submitted for publication). The ALKs are more similar with each other than with ActR-II and -IIB, T β R-II, and Daf-1, and thus, they form a subfamily among the putative serine/threonine kinase receptors. Ligands for ALKs remain to be determined. A recent report showed that Tak 7L, a mouse counterpart of human ALK-2, binds TGF- β 1 and forms a cross-linked complex with a size similar to that of the TGF- β type I receptor (17). In this communication, we report the cloning of another serine/threonine kinase receptor, termed ALK-6, from a mouse embryo cDNA library. Interestingly, mouse ALK-6 is structurally very similar to human and mouse ALK-3, but their expression patterns are very different.

MATERIALS AND METHODS

Cloning of cDNAs for Mouse ALK-3 and ALK-6. As a probe for screening a cDNA library, a PCR recombinant 11.1 (16), which corresponds to the kinase domain of human ALK-4, was used. A 12 day mouse embryo λ EX10x cDNA library (Novagen, Madison, WI) was screened with the 11.1 probe labeled by the Megaprime labeling system (Amersham, U.K.). Hybridization to nitrocellulose replica filters was performed as described previously (18). The filters were washed two times with 2 x SSC (1 x SSC is 15 mM sodium citrate, 150 mM NaCl) and 0.1% SDS at 37°C for 15 min, and two times with 0.5 x SSC and 0.1% SDS at 55°C for 15 min. Purification of bacteriophages were performed as previously described (18). Twenty-four positive clones were obtained. They were analyzed by Southern blot hybridization using specific probes for ALK-1 to -4. In order to obtain the 5' part of mouse ALK-3, the cDNA library was screened with a probe corresponding to nucleotides 79-824 of human ALK-3 (16). Nucleotide sequencing was performed on both strands (19) using Sequenase (U.S. Biochemical Corporation, Cleveland, OH) and specific oligonucleotide primers. Compressions were resolved using 7-deaza-GTP (U.S. Biochemicals). DNA sequences were analyzed by DNA STAR computer program (DNA STAR, Ltd. U.K.).

Southern Blot Hybridization. DNAs from positive clones were digested with *Eco*RI and *Hind*III, electrophoretically separated on a 1.3% agarose gel and transferred to nitrocellulose filters as described (20). The filters were then hybridized with specific probes (16) for human ALK-1 (nucleotides 288-670), ALK-2 (nucleotides 1-581), ALK-3 (nucleotides 79-824) or ALK-4 (nucleotides 1178-1967).

Northern Blot Hybridization. A multiple mouse tissue blot was obtained from Clontech (Palo Alto, CA). The filter was hybridized with probes for mouse ALK-3 and ALK-6. The *Eco*RI - *Pst*I restriction fragment, corresponding to nucleotides 79-1100 of ALK-3, and the *Sac*I - *Hpa*I fragment, corresponding to nucleotide 57-720 of ALK-6, were used as probes. Hybridization was performed with ³²P-labeled probes at 42°C overnight in 50% formamide, 5 x SSC, 0.1% SDS, 50 mM sodium phosphate, pH 7.0, 5 x Denhardt's solution, and 0.1 mg/ml salmon sperm DNA. The filter was washed at 65°C, two times for 30 min in 2.5 x SSC, 0.1% SDS, and two times for 30 min with 0.3 x SSC, 0.1% SDS. The filter was then subjected to autoradiography. Stripping of the blot was performed by incubation in distilled water at 90-100°C for 20 min.

RESULTS AND DISCUSSION

Cloning of Mouse ALK-6 cDNA. By screening the 12 day mouse embryo cDNA library using a probe from the kinase domain of ALK-4 under low stringency hybridization conditions, we obtained 20 positive clones. DNAs from these clones were analyzed by Southern blot hybridization in order to investigate whether they were the mouse homologs of human ALK-1 to -4. Seven clones hybridized very strongly with the ALK-2 probe, seven clones hybridized with the ALK-3 probe, and two clones hybridized with the ALK-4 probe. None of them hybridized to ALK-1 specific probes. Four clones hybridized to the probe corresponding to the conserved kinase domain of ALK-4, but not to probes from more divergent parts of ALK-1 to -4. Analysis of these clones revealed that they have an identical sequence, which was different from those of ALK-1 to -5; therefore, the novel clone was termed ALK-6. The longest clone ME 6 with a 2.0 kb insert was completely sequenced.

Sequence of Mouse ALK-6. Sequencing of ME 6 yielded a 1952 bp fragment consisting of an open reading frame of 1509 bp (503 amino acids), flanked by a 5' untranslated sequence of 186 bp, and a 3' untranslated sequence of 157 bp. The nucleotide and predicted amino acid sequences are shown in Fig. 1. No polyadenylation signal was found in the 3' untranslated region of ME 6, indicating that the cDNA was internally primed in the 3' end. Only one ATG codon was found in the 5' part of the open reading frame, which fulfilled the rules for translation initiation (21), and was preceded by an in-frame stop codon at nucleotides 163-165. However, a typical hydrophobic leader sequence was not observed at the N-terminus of the translated region. Since there is no ATG codon and putative hydrophobic leader sequence, this ATG codon is likely to be used as a translation initiation site. It remains to be determined how efficient the mouse ALK-6 is transported to the cell membrane. Signal peptidase possibly cuts between amino acids 13 and 14, which is a preferable site according to the von Heijne algorithm (22).

Similar to other ALKs, ALK-6 has a relatively short extracellular domain, followed by a transmembrane domain, and an intracellular putative serine/threonine kinase domain. The extracellular domain has a cysteine-rich domain, which is likely to bind ligands. The serine/threonine kinase receptors, thus far reported, have one or more potential N-glycosylation sites (11-16). However, the mouse ALK-6 does not have any potential N-glycosylation sites.

The intracellular domain of ALK-6 consists almost entirely of a putative kinase domain. A consensus sequence for the binding of ATP (Gly-X-Gly-X-X-Gly in subdomain I, followed by a lysine residue further downstream in subdomain II;

nomenclature according to Ref. 23) is found. Analysis of the amino acid sequences in subdomains VI and VIII, which are most useful to predict the specificity of amino acid phosphorylation (23), indicates that ALK-6 is a serine/threonine kinase. Two kinase inserts were observed between subdomains VIA and VIB, and between subdomains X and XI. The amino acid sequence of mouse ALK-6 is most similar to human ALK-3 among the serine/threonine kinase receptors (16). To rule out the possibility that ALK-6 is the mouse counterpart of ALK-3, we cloned a mouse cDNA for ALK-3 and compared it with the mouse ALK-6.

Cloning of Mouse ALK-3 cDNA. Southern blot analysis revealed that a clone termed ME-7 hybridized with the human ALK-3 probe, however, nucleotide sequencing revealed that this clone was incomplete, and lacked the 5' part of the translated region. Therefore, the same cDNA library was screened by a probe corresponding to the extracellular domain of human ALK-3, and one positive clone, ME-D, was obtained. The clone was isolated and the sequence was analyzed. Although ME-D was incomplete in the 3' end of the translated region, ME-7 and ME-D overlapped and together covered the complete sequence of mouse ALK-3. The predicted amino acid sequence of mouse ALK-3 is very similar to the human sequence; only 8 amino acid residues differ (98% identity) (Fig. 2).

Comparison of Mouse ALK-3 and ALK-6. The amino acid sequences of mouse ALK-3 and -6 are 71% identical; the identities in the kinase domains and the cysteine-rich putative ligand-binding domains are 85% and 59%, respectively. The amino acid sequence similarities in the extracellular cysteine-rich domains of other ALKs and ActR-II, T β R-II and Daf-1 are lower (less than 40%). High sequence similarity in this domain has been reported for ActR-II and ActR-IIB (64% identity), which bind the same ligand. Thus, ALK-3 and ALK-6 are closely related molecules in the serine/threonine kinase receptor family, and may bind the same or closely related ligand(s). The calculated molecular weights of the primary translated products of mouse ALK-3 and -6, without the putative signal sequences, are 57,447 and 55,576, respectively. Therefore, the molecular weights are closer to those of type I receptors than type II receptors. The intracellular portions of ALK-3 and -6 are highly conserved compared to the extracellular domains. However, the intracellular juxtamembrane domain and at the kinase insert between subdomains X and XI show relatively high divergence. Whether these portions functions, e.g. in the association with downstream components in the signal transduction pathway, remains to be elucidated.

Expression of mRNA for Mouse ALK-3 and ALK-6. The distribution of mRNA for mouse ALK-3 and -6 in various mouse tissues was determined by Northern

blot analysis (Fig. 3). In order to avoid cross-hybridization, probes corresponding to the 5' untranslated and extracellular regions were used. Using the probe for mouse ALK-3, a 1.1 kb transcript was found only in spleen. By reprobing the blot with the ALK-6 specific probe, a transcript with 7.2 kb was found in brain. A weak band was also seen in lung; however, bands were not seen in the other tissues tested. Most of the serine/threonine kinase receptors are widely expressed, exemplified by ALK-2, and -4, and TBR-II (14, 16). In contrast, the expression profile of human ALK-3 is relatively restricted, and fragments of 4.4 and 7.9 kb were found most abundantly in skeletal muscle (16). Expression in spleen was not tested for the human ALK-3. The transcript size for mouse ALK-3 is smaller than that of the cloned cDNA. Larger size transcripts may possibly be seen in other tissues which were not tested in the present study, and/or at other developmental stages. As the ALK-3 coding region is 1596 bp, the 1.1 kb transcript is possibly derived from a differently spliced mRNA. The functional significance remains to be elucidated. Difference in mRNA splicing in the region coding for the extracellular domain may lead to the production of soluble binding protein. Difference in the intracellular domain may lead to a truncated cell surface receptor that possibly acts in a dominant negative fashion. Different expression profiles between ALK-3 and -6 suggest different roles of these receptors *in vivo*. Growth regulatory factors have been shown to act in autocrine and/or paracrine fashion. If this is also the case for ALK-3 and -6, they may bind proteins with restricted expression profiles. For example, certain members in the BMP family were shown to be produced with highly limited expression patterns (24-26). Further investigation are in progress in order to identify the ligand(s) for ALK-3 and -6.

Acknowledgments: We thank Kari Ahtalo at Helsinki University, Finland for the cDNA library. We also thank Christer Wernstedt for preparing oligonucleotides. Peter ten Dijke is supported by an EMBO fellowship. Nucleotide sequences of mouse ALK-3 and -6 are deposited in GenBank.

REFERENCES

1. Roberts, A.B. and Sporn, M.B. (1990) In *Peptide Growth Factors and Their Receptors, part I* (M.B. Sporn and A.B. Roberts, eds.) pp. 419-472, Springer-Verlag, Berlin.
2. Vale, W., Hauch, A., Rivier, C. and Yu, J. (1990) In *Peptide Growth Factors and Their Receptors, part II* (M.B. Sporn and A.B. Roberts, eds.) pp. 211-248, Springer-Verlag, Berlin.
3. Lyons, K.M., Jones, C.M. and Hogan, B.L.M. (1991) *Trends Genet.* 7, 408-412.
4. Massagué, J. (1992) *Cell* 69, 1067-1070.
5. Lin, H.Y. and Lodish, H.F. (1993) *Trends Cell Biol.* 3, 14-19.
6. Laiho, M., Wells, F.M.B. and Massagué, J. (1990) *J. Biol. Chem.* 265, 18518-18524.

7. Laiho, M., Wels, F.M.B., Boyd, F.T., Ignatz, R.A. and Massagué, J. (1991) *J. Biol. Chem.* 266, 9108-9112.
8. Wrana, J.L., Attisano, L., Cárcamo, J., Zentella, A., Doody, J., Laiho, M., Wang, X.-F. and Massagué, J. (1992) *Cell* 71, 1003-1014.
9. Hino, M., Tojo, A., Miyazono, K., Miura, Y., Chiba, S., Eto, Y., Shibai, H. and Takaku, F. (1989) *J. Biol. Chem.* 264, 10309-10314.
10. Paralkar, V. M., Hammonds, R.G. and Reddi, A.H. (1991) *Proc. Natl. Acad. Sci. USA* 88, 3397-3401.
11. Mathews, L.S. and Vale, W.W. (1991) *Cell* 65, 973-982.
12. Attisano, L., Wrana, J.L., Cheifetz, S. and Massagué, J. (1992) *Cell* 68, 97-108.
13. Mathews, L.S., Vale, W.W. and Kintner, C.R. (1992) *Science* 255, 1702-1705.
14. Lin, H.Y., Wang, X.-F., Ng-Eaton, E., Weinberg, R.A. and Lodish, H.F. (1992) *Cell* 68, 775-785.
15. Georgi, L.L., Albert, P.S. and Riddle, D.L. (1990) *Cell* 61, 635-645.
16. ten Dijke, P., Ichijo, H., Franzén, P., Schulz, P., Sarau, J., Toyoshima, H., Heldin, C.-H. and Miyazono, K. (1993) *Oncogene* in press.
17. Ebner, R., Chen, R.-H., Shum, L., Lawler, S., Zloncheck, T.F., Lee, A., Lopez, A.R. and Derynck, R. (1993) *Science* 260, 1344-1348.
18. Kanzaki, T., Olofsson, A., Morén, A., Wernstedt, C., Hellman, U., Miyazono, K., Claesson-Welsh, L. and Heldin, C.-H. (1990) *Cell* 61, 1051-1061.
19. Sanger, F., Nicklen, S. and Coulson, A.R. (1977) *Proc. Natl. Acad. Sci. USA* 74, 3463-3467.
20. Sambrook, J., Fritsch, E.F. and Maniatis, T.E. (1989) *Molecular Cloning: A Laboratory Manual*, 2nd ed., pp. 9.32-9.62, Cold Spring Harbor Laboratory, NY.
21. Kozak, M. (1987) *Nucl. Acids Res.* 15, 8125-8148.
22. von Heijne, G. (1986) *Nucl. Acids Res.* 14, 4683-4690.
23. Hanks, S.K., Quinn, A.M. and Hunter, T. (1988) *Science* 241, 42-52.
24. Lee, S.-J. (1991) *Proc. Natl. Acad. Sci. USA* 88, 4250-4254.
25. McPherron, A.C. and Lee, S.-J. (1993) *J. Biol. Chem.* 268, 3444-3449.
26. Özkaynak, E., Schneegelsberg, P.N.J., Jin, D.F., Clifford, G.M., Warren, F.D., Drier, E.A. and Oppermann, H. (1992) *J. Biol. Chem.* 267, 25220-25227.

Figure Legends

Fig. 1. Nucleotide and deduced amino acid sequences of mouse ALK-6. The putative signal sequence and transmembrane domain are overlined over the amino acid sequence. Cysteine residues found in the extracellular domain are boxed. The borders of the kinase domain are marked by arrows. The stop codon which ends the open reading frame is shown by an asterisk. The in-frame stop codon is underlined.

Fig. 2. Comparison of the amino acid sequences of human and mouse ALK-3 and mouse ALK-6. Identical amino acids are boxed and conserved cysteine residues in the extracellular domains are shaded. Potential N-glycosylation sites for human and mouse ALK-3 are shown by a thick line. Putative cleavage sites by signal peptidase are indicated by Δ . Putative transmembrane domains for human and mouse ALK-3 are double overlined. The borders of the kinase domains are indicated by arrows. Kinase subdomains (23) are indicated by roman numerals. The two kinase inserts are underlined (thin lines).

Fig. 3. Northern blot analysis of mouse ALK-3 and -6 mRNA expression in different mouse tissues. A multiple mouse tissue blot (Clontech) was hybridized with probes for mouse ALK-3 (A) and ALK-6 (B). Size markers are indicated on the right. Specific bands are indicated by arrows on the left.

Fig. 1

human ALK-3 1
mouse ALK-3 1
mouse ALK-6 1

MIDLYIVIRLLGAYEFTISRVGGQMLDSHLNGTGNKSDLSOKKSENGVTLAPEDTLPLK
V

NLRS3GKLNVCITKEDGESIAPSIRPKI--LR

human ALK-3 61
mouse ALK-3 61
mouse ALK-6 32

YSGHPPDQAINNTITNGHFAIIEEDDQGETTLASGHRKYEGSDFOKDSPKAQLR
YSGHPPDQAINNTITNGHFAIIEEDDQGETTLASGHRKYEGSDFOKDSPKAQLR
KHHHHPEDSVKNTISTDGVETMTIEEDDSCHPVVLSGELGLGSSDFQDRDTPHQR

human ALK-3 121
mouse ALK-3 121
mouse ALK-6 92

TIEHNTMLNDYLOPTLPVVGPFPOGSTRVLLTISHAVCTIAHIFSSCFCKHY
TIEHNTMLNDYLOPTLPVVGPFPOGSTRVLLTISHAVCTIAHIFSSCFCKHY
SLEHNTMLNDYLOPTLPVVGPFPOGSTRVLLTISHAVCTIAHIFSSCFCKHY
ITERHNEHKKDLPPLPKDRDEVIDGPIHHKALLLISVTVCSLLLVLLIL-ECYFR

human ALK-3 160
mouse ALK-3 180
mouse ALK-6 151

CKSTSSRRRYNRDLEQDEAFIPVGESLKKOLLQDSOSSGSSGSLPLLVORTIAKQIDHVRQ
CKSTSSRRRYNRDLEQDEAFIPVGESLKKOLLQDSOSSGSSGSLPLLVORTIAKQIDHVRQ
-KROEARPRYSICLEDDEFTVLPFGESLRLDLEDSOSSGSSGSLPLLVORTIAKQIDHVRQ

human ALK-3 240
mouse ALK-3 240
mouse ALK-6 210

VGGGRNYGEVWNGKVRGGERVAVKVFITTEASVFRETEITYOTVLRHRENILCFIAADIKCT
VGGGRNYGEVWNGKVRGGERVAVKVFITTEASVFRETEITYOTVLRHRENILCFIAADIKCT
IGKGRYGEVWNGKVRGGERVAVKVFITTEASVFRETEITYOTVLRHRENILCFIAADIKCT

I

II

III

IV

human ALK-3 300
mouse ALK-3 300
mouse ALK-6 270

GSVTOLYLLTTOYHENGSLYDFLKCAITLQTRALLKLAYSAAACGLCHLHTEITYGTQCKPAIA
GSVTOLYLLTTOYHENGSLYDFLKCAITLQTRALLKLAYSAAACGLCHLHTEITYGTQCKPAIA
GSVTOLYLLTTOYHENGSLYDFLKCAITLQTRALLKLAYSAAACGLCHLHTEITYGTQCKPAIA

V

VIA

human ALK-3 360
mouse ALK-3 360
mouse ALK-6 330

MRDLKSKNLLIKKNGSCCIAOLGLAVKFNSTQNEVDVPLNTRVGTKRYNAPEVLDESINR
MRDLKSKNLLIKKNGSCCIAOLGLAVKFNSTQNEVDVPLNTRVGTKRYNAPEVLDESINR
MRDLKSKNLLIKKNGSCCIAOLGLAVKFNSTQNEVDVPLNTRVGTKRYNAPEVLDESINR

VIB

VII

VIII

human ALK-3 420
mouse ALK-3 420
mouse ALK-6 390

MHFOPIYINADITVSFGITIVENARRCTITGGIVEEYQLPYYNNHVPSPSYEONREVVCKRL
MHFOPIYINADITVSFGITIVENARRCTITGGIVEEYQLPYYNNHVPSPSYEONREVVCKRL
MHFOPIYINADITVSFGITIVENARRCTITGGIVEEYQLPYYNNHVPSPSYEONREVVCKRL

IX

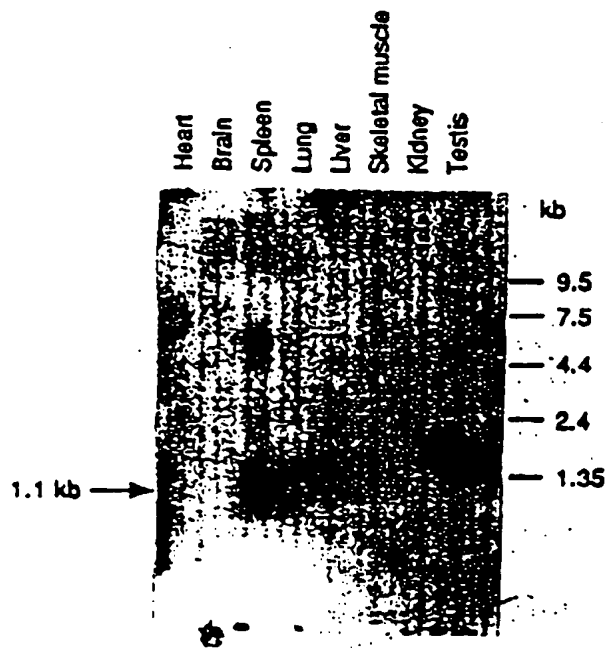
X

human ALK-3 480
mouse ALK-3 480
mouse ALK-6 450

RPIVSNRNVNSDECTKAVLKLHSECVAAHMPASRLTALRIKKTILAKHVESQDVAKI
RPIVSNRNVNSDECTKAVLKLHSECVAAHMPASRLTALRIKKTILAKHVESQDVAKI
RPIVSNRNVNSDECTKAVLKLHSECVAAHMPASRLTALRIKKTILAKHVESQDVAKI

XI

(A) Mouse ALK-3



(B) Mouse ALK-6

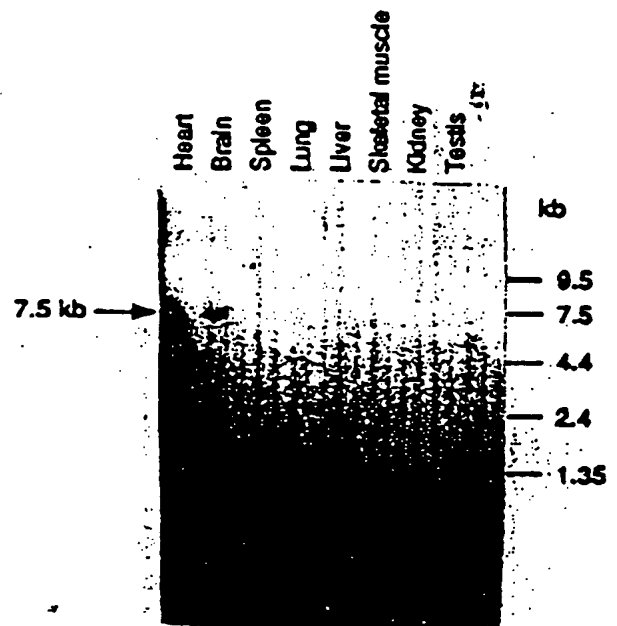


Fig. 3

A. 4 final seq Map (1 > 2233)

Enzymes : All 206 enzymes (No Filter)

Site and Sequence

Settings:

Linear, Certain & Uncertain Sites, Standard Genetic Code

ATGGCGGAGTCGGCCGGAGCCCTCCTCCTTCTTCCCTTGTGTGTCTCCTGCTCGCCGGCAGCGCGGGT 70
A E S A G A S S F F P L V V L L L A G S G G

GGCCCCCGGGGGTCCAGGCTCTGCTGTGTGCGTGCACCAGCTGCCTCCAGGCCAACTACACGTGTGA 140
S G P R G V Q A L L C A C T S C L Q A N Y T C E

GACAGATGGGGCTGCATGCTTTCCATTTTCAATCTGGATGGGATGGAGCACCATGTGCGCACCTGCATC 210
T D G A C H V S I F N L D G H E H H V R T C I

CCCAAAGTGGAGCTGGTCCCTGCCGGGAAGCCCTTCTACTGCCTGAGCTCGGAGGACCTGCGCAACACCC 280
P K V E L V P A G K P F Y C L S S E D L R N T

ACTGCTGCTACACTGACTACTGCAACAGGATCGACTTGAGGGTGGCCAGTGGTCACCTCAAGGAGCCTGA 350
H C C Y T D Y C N R I D L R V P S Q H L K E P E

GCACCCGTCATGTGGGGCCCCGTGGAGCTGGTAGGCATCATCGCCGGCCCGGTGTTCTCTCTGTTCTC 420
H P S H W G P V E L V G I I A G P V F L L F L

ATCATCATCATTGTTTTCTTGTCTTAATACTATCATCAGCGTGTCTATCACAACCGCCAGAGACTGGACA 490
I I I I V F L V I N Y H Q R V Y H N R Q R L D

TGGAAGATCCCTCATGTGAGATGTGTCTCTCCAAAGACAAGACGCTCCAGGATCTTGTCTACGATCTCTC 560
H E D P S C E M C L S K D K T L Q D L V Y D L S

CACCTCAGGGTCTGGCTCAGGGTTACCCCTCTTTGTCCAGCGCACAGTGGCCCGAACCATCGTTTTACAA 630
T S G S G S G L P L F V Q R T V A R T I V L Q

GAGATTATTGGCAAGCGCTCGGTTTGGGGAAGTATGGCGGGGGCCCTGCAGGGGTGGTGATGTGGCTGTGA 700
E I I G K G R F G E V W R G R W R G G D V A V

AAATATTCTTTCTCGTGAAGAACGGTCTTGGTTTCAGGGAAGCAGAGATATACCAGACGGTCTGCTGCG 770
K I F S S R E E R S W F R E A E I Y Q T V M L R

CCATGAAAACATCCTTGGATTATTGCTGCTGACAATAAAGATAATGGCACCTGGACACAGCTGTGGCTT 840
H E N I L G F I A A D N K D N G T W T Q L W L

GTTTCTGACTATCATGAGCAGGGTCCCTGTTTGATTATCTGAACCGGTACACAGTGACAATTGAGGGGA 910
V S D Y H E H G S L F D Y L N R Y T V T I E G

TGATTAAGCTGGCCTTGTCTGCTGCTAGTGGGCTGGCACACCTGCACATGGAGATCGTGGGCACCCAAGG 980
M I K L A L S A A S G L A H L H M E I V G T Q G

GAAGCCTGGAATTGCTCATCGAGACTTAAAGTCAAAGAACATTCTGGTGAAGAAAAATGGCATGTGTGCC 1050
K P G I A H R D L K S K N I L V K K N G M C A

ATAGCAGACCTGGGCCTGGCTGTCCGTCATGATGCAGTCACTGACACCATTGACATTGCCCCGAATCAGA 1120
I A D L G L A V R H D A V T D T I D I A P N Q

GGGTGGGGACCAAACGATACATGGCCCCCTGAAGTACTTGATGAAACCATTAATATGAAACACTTTGACTC 1190
R V G T K R Y H A P E V L D E T I N M K H F D S

CTTTAAATGTCTGATATTTATGCCCTCGGGCTTGATATTGGGAGATTGCTCGAAGATGCAATTCTGGA 1260
F K C A D I Y A L G L V Y W E I A R R C N S G

GGAGTCCATGAAGAATATCAGCTGCCATATTACGACTTAGTGCCCTCTGACCCCTTCATTGAGGAAATGC 1330
G V H E E Y Q L P Y Y D L V P S D P S I E E H

GAAAGGTTGTATGTGATCAGAAGCTGCGTCCCAACATCCCCAACTGGTGGCAGAGTTATGAGGCACTGCG 1400
R K V V C D O K L R P N I P N W W Q S Y E A L R

GGTGATGGGCAAGATGATGCCAGAGTGTGGTATGCCAACGGCGCAGCCCGCCTGACGGCCCTGCGCATC 1470
V H G K M H R E C W Y A N G A A R L T A L R I

Site and Sequence

CCCCCTCCAGGTCAGCGTGCAGGAAGACGTGAAGATCTAACTGCTCCCTCTCTCCACACGGA 1840

T L S Q L S V Q E D V K I .

GTCTCTGGCAGCGAGAACTACGCACAGCTGCCGCGTTGAGCGTACGATGGAGGCCCTACCTCTCGTTTCTG	1810
CCCAGCCCTCTGTGGCCAGGAGCCCTGGCCCCGAAGAGGGACAGAGCCCGGGAGAGACTCGCTCACTCCC	1880
ATGTTGGGTTTGAGACAGACACCTTTTCTATTTACCTCCTAATGGCATGGAGACTCTGAGAGCGAATTGT	1750
GTGGAGAACTCAGTCCACACCTCGAACTCGTTGTAGTGGGAAGTCCCCGGAAACCCGGTGCACTCGGCA	1820
CGTGGCCAGGAGCCATGACAGGGGCGCTTGGGAGGGGCCGGAGGAACCGAGGTGTTGCCAGTGCTAAGCT	1890
GCCCTGAGGGTTTCTTCGGGGACCAGCCACAGCACACCAAGGTGGCCCGGAAGAACCAGAAGTGCAGC	1960
CCCTCTCACAGGCAGCTCTGAGCCGCGCTTTCCCTCTCCTCGGATGGACGCTGCCGGGAGACTGCCA	2030
GTGGAGACGGAATCTGCCGCTTTGTCTGTCCAGCCGTGTGTGCATGTGCCGAGGTGCGTCCCCCGTTGTG	2100
CCTGGTTCTGCCATGCCCTTACACGTGCGTGTGAGTGTGTGTGTGTCTGTAGGTGCGGCACTTACCTG	2170
CTTGAGCTTTCTGTGCATGTGCAGGTGGGGGTGTGGTGTGTATGCTGTCCGTGCTTGCTGGTGCCTCTT	2240
TTCAGTAGTGAGCAGCATCTAGTTTCCCTGGTGGCCTTCCCTGGAGGTGTCTCCCTCCCCCAGAGCCCTT	2310
CATGCCACAGTGGTACTCTGTGT	2333